



Discover Generics

Cost-Effective CT & MRI Contrast Agents



WATCH VIDEO

AJNR

Imaging Patterns Characterizing Mitochondrial Leukodystrophies








S.D. Roosendaal, T. van de Brug, C.A.P.F. Alves, S. Blaser, A. Vanderver, N.I. Wolf and M.S. van der Knaap

AJNR Am J Neuroradiol published online 1 April 2021

<http://www.ajnr.org/content/early/2021/04/15/ajnr.A7097>

This information is current as of June 18, 2025.

Imaging Patterns Characterizing Mitochondrial Leukodystrophies

 S.D. Roosendaal,  T. van de Brug,  C.A.P.F. Alves,  S. Blaser,  A. Vanderver,  N.I. Wolf, and  M.S. van der Knaap



ABSTRACT

BACKGROUND AND PURPOSE: Achieving a specific diagnosis in leukodystrophies is often difficult due to clinical and genetic heterogeneity. Mitochondrial defects cause 5%–10% of leukodystrophies. Our objective was to define MR imaging features commonly shared by mitochondrial leukodystrophies and to distinguish MR imaging patterns related to specific genetic defects.

MATERIALS AND METHODS: One hundred thirty-two patients with a mitochondrial leukodystrophy with known genetic defects were identified in the data base of the Amsterdam Leukodystrophy Center. Numerous anatomic structures were systematically assessed on brain MR imaging. Additionally, lesion characteristics were scored. Statistical group analysis was performed for 57 MR imaging features by hierarchic testing on clustered genetic subgroups.

RESULTS: MR imaging features indicative of mitochondrial disease that were frequently found included white matter rarefaction ($n = 50$ patients), well-delineated cysts ($n = 20$ patients), T2 hyperintensity of the middle blade of the corpus callosum ($n = 85$ patients), and symmetric abnormalities in deep gray matter structures ($n = 42$ patients). Several disorders or clusters of disorders had characteristic features. The combination of T2 hyperintensity in the brain stem, middle cerebellar peduncles, and thalami was associated with complex 2 deficiency. Predominantly periventricular localization of T2 hyperintensities and cystic lesions with a distinct border was associated with defects in complexes 3 and 4. T2-hyperintense signal of the cerebellar cortex was specifically associated with variants in the gene *NUBPL*. T2 hyperintensities predominantly affecting the directly subcortical cerebral white matter, globus pallidus, and substantia nigra were associated with Kearns-Sayre syndrome.

CONCLUSIONS: In a large group of patients with a mitochondrial leukodystrophy, general MR imaging features suggestive of mitochondrial disease were found. Additionally, we identified several MR imaging patterns correlating with specific genotypes. Recognition of these patterns facilitates the diagnosis in future patients.

ABBREVIATIONS: KSS = Kearns-Sayre syndrome; SDH = Succinate dehydrogenase; MELAS = mitochondrial encephalopathy, lactic acidosis, and stroke-like episodes syndrome; MNGIE = mitochondrial neurogastrointestinal encephalopathy

Leukodystrophies are genetic disorders predominantly involving the white matter of the CNS.¹ Clinical and genetic heterogeneity complicate establishing a specific molecular diagnosis, which becomes more effective when clinical, radiologic, and genetic parameters are combined.¹ Most leukodystrophies are

characterized by a specific combination of affected structures on MR imaging, constituting a recognizable pattern. Recent diagnostic algorithms for leukodystrophies acknowledge the central role of MR imaging pattern recognition.^{1,2}

A subset of leukodystrophies, involving 5%–10%, is caused by mitochondrial defects.^{3,4} Mitochondrial diseases comprise a group of disorders with genetic defects in mitochondrial oxidative energy metabolism. Mitochondria are cellular organelles that produce energy via oxidative phosphorylation (Online Supplemental Data). The oxidative phosphorylation system consists of 5 respiratory

Received November 17, 2020; accepted after revision January 14, 2021.

From the Departments of Radiology (S.D.R.) and Epidemiology and Biostatistics (T.v.d.B.), Amsterdam UMC, Amsterdam, the Netherlands; Division of Neuroradiology (C.A.P.F.A.), and Department of Radiology and Division of Neurology (A.V.), The Children's Hospital of Philadelphia, Philadelphia, Pennsylvania; Division of Neuroradiology (S.B.), Department of Diagnostic Imaging, Hospital for Sick Children, University of Toronto, Toronto, Ontario, Canada; Department of Pediatric Neurology (M.S.v.d.K., N.I.W.), Emma Children's Hospital, Amsterdam Leukodystrophy Center, Amsterdam UMC, Vrije Universiteit and Amsterdam Neuroscience, Amsterdam, the Netherlands; and Department of Functional Genomics (M.S.v.d.K.), Center for Neurogenomics and Cognitive Research, VU University, Amsterdam, the Netherlands.

The following authors are members of the European Reference Network for Rare Neurological Disorders, project ID 739510: M.S. van der Knaap and N.I. Wolf.

Please address correspondence to Stefan D. Roosendaal, MD, PhD, Amsterdam UMC, Department of Radiology, Meibergdreef 9, 1105 AZ Amsterdam, the Netherlands; e-mail: stefan.roosendaal@amsterdamumc.nl

 Indicates open access to non-subscribers at www.ajnr.org

 Indicates article with online supplemental data.

<http://dx.doi.org/10.3174/ajnr.A7097>

chain complexes in the mitochondrial membrane. Human cells possess 2 different genomes: the nuclear genome, containing approximately 20,000 genes, and the mitochondrial genome, containing 37 genes. Although mitochondria have functions in addition to energy metabolism, mitochondrial diseases are typically regarded as disorders of oxidative phosphorylation.⁵ They are caused by variants in either mitochondrial genes or, much more frequently, nuclear genes encoding proteins necessary for mitochondrial function.⁵ Concerning the high number of associated gene defects, it is not surprising that mitochondrial disorders have a striking clinical heterogeneity;⁵ the high energy demand of the CNS explains it being the most commonly affected organ.⁵

Some mitochondrial encephalopathies predominantly manifest with lesions of gray matter structures (eg, mitochondrial encephalopathy, lactic acidosis, and stroke-like episodes syndrome (MELAS); Alpers syndrome; and Leigh syndrome).⁶ Another category manifests as leukodystrophy.⁷ The leukoencephalopathy in Kearns-Sayre syndrome (KSS)⁸ and mitochondrial neurogastrointestinal encephalopathy syndrome (MNGIE)⁹ has been known for a long time, but more recently, next-generation genetic sequencing techniques have revealed that numerous unsolved cases of leukodystrophy have a mitochondrial cause, and MR imaging patterns of new mitochondrial leukodystrophies were added.¹⁰ Patient selection by specific MR imaging patterns was pivotal in the detection of the novel leukodystrophies caused by defects in the genes *LYRM7*, *APOPT1*, *NUBPL*, and *CLPP*.^{11–14} However, systematic studies on MR imaging patterns in larger groups of patients with different mitochondrial leukodystrophies are lacking, and information on the associated MR imaging patterns is lacking in recent leukodystrophy diagnostic algorithms.^{1,2}

In the current study, we systematically analyzed MRIs of a large group of patients with mitochondrial leukodystrophy. In all patients, the causative gene defect was known. The study aims were to describe MR imaging features suggesting a mitochondrial leukodystrophy in general and to distinguish MR imaging patterns related to particular gene defects.

MATERIALS AND METHODS

Patients

In the MR imaging data base of the Amsterdam Leukodystrophy Center, comprising more than 4000 patients, 132 patients with a mitochondrial leukodystrophy and a known gene defect were identified. Patients with defects in genes encoding mitochondrial transfer RNA synthetases were excluded because the MR imaging patterns associated with the respective disorders have been well-described in separate studies and their high numbers in the data base would dominate the current study.^{15–18} The retrospective phenotype analysis study was approved by the Ethics Committee of the VU University Medical Center.

Of the 132 patients, 43 had a respiratory chain complex 1 defect: 22 with variants in the genes *NDUFV1*, 1 in *NDUFV2*, 10 in *NDUFS1*, 2 in *NDUFS3*, 2 in *NDUFS4*, 1 in *NDUFS6*, 1 in *NDUFS7*, 3 in *NDUFS8*, and 1 in *NDUFA5*. Complex 2 deficiency was present in 22 patients: 4 with variants in *SDHA*, 5 in *SDHB*, and 10 in *SDHAF1*; in 3 patients with biochemical evidence of complex 2 deficiency, the gene defect was not determined, and in one of them the *SDHA* protein expression was reduced. Most of these patients were part of a previous study on complex 2 deficiency.¹⁹ Complex 3

deficiency was present in 10 patients, all with variants in *LYRM7*. Some were part of a previous study on *LYRM7*.¹⁴ Complex 4 deficiency was present in 9 patients: 7 with variants in *APOPT1*, 1 with variants in *COX6B*, and 1 with variants in *COX10*. Most patients with *APOPT1* variants were part of a previous study on *APOPT1*.¹² Iron-sulfur cluster assembly pathway variants were present in 33 patients: 9 with variants in *NUBPL*, 8 in *IBA57*, 7 in *ISCA2*, 5 in *FDX1L*, 3 in *BOLA3*, and 1 in *CDK5RAP1*. Descriptions of some of the patients with *NUBPL*,¹¹ *IBA57*,²⁰ and *FDX1L*²¹ variants were published before. A large mitochondrial DNA deletion was present in 6 patients, all presenting with the KSS phenotype. Additionally, 4 patients had *TYMP* variants, indicative of MNGIE, and 5 patients had *CLPP* variants; descriptions of some of the latter have been published before.¹³

MR Imaging Evaluation

At least 1 brain MR imaging was available for each patient. When multiple MRIs were available, the first was used for analysis. MR sequences and image quality were variable because the MRIs were acquired at multiple institutions. T2-weighted and nonenhanced T1-weighted sequences were available for every patient; FLAIR was available for 115 patients. These images were used to systematically score numerous white and gray matter structures as normal or abnormal, and specific features of the abnormalities were scored, as described in van der Knaap et al.²² More salient features of this scoring system were as follows: The white matter was divided into 3 zones—directly subcortical white matter (ie, arcuate fibers), deep white matter, and periventricular white matter. The corpus callosum was divided into inner, middle, and outer blades (or layers) and anterior, middle, and posterior parts. Cysts were strictly scored as lesions with a signal intensity identical to that of CSF on FLAIR images, whereas lesions having a signal intensity on FLAIR intermediate between normal white matter and CSF were defined as rarefaction. The caudate nucleus, putamen, globus pallidus, thalamus, dentate nucleus, substantia nigra, cerebral cortex, and cerebellar cortex were individually evaluated.

Statistical Analysis

Group analysis was performed with 57 MR imaging features. We first analyzed which abnormalities were most commonly observed among our 132 patients.

Considering the large number of different genetic subgroups, it was unlikely that specific patterns would survive multiple comparison analysis. We therefore clustered genetic subgroups. First, patients were clustered in 9 groups: complex 1; complex 2; complex 3; complex 4; iron-sulfur cluster defects excluding *NUBPL*; *NUBPL*; MNGIE; KSS; and *CLPP*. This clustering was partly on the basis of the type of mitochondrial defect and partly on the basis of prior knowledge from personal experience and reviewing the MRIs. To further reduce the number of subgroups, we visually inspected a heat map of the data, in which the patients were ordered by genetic subtype and the features were clustered using unsupervised hierarchical cluster analysis. No differences were observed between defects of complexes 3 and 4, and they were clustered. *NUBPL* and KSS were considered separately. Complex 1 defects were clustered

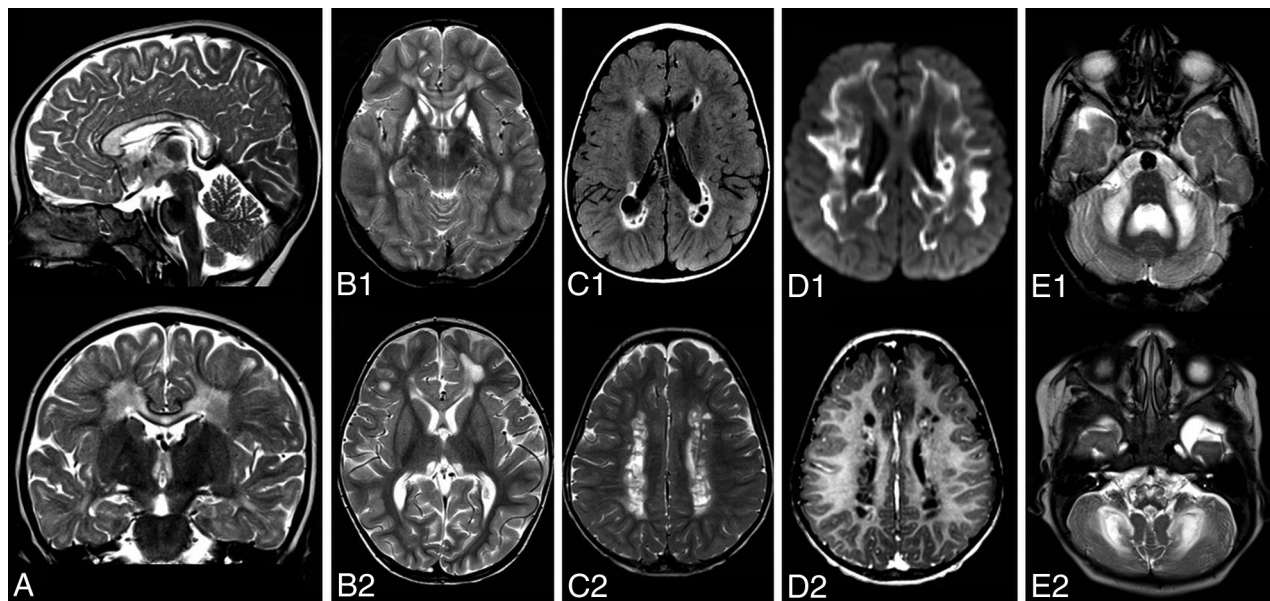


FIG 1. MR imaging features suggestive of mitochondrial leukodystrophy. A, Sagittal and coronal T2-weighted images of a patient with a complex 2 deficiency show longitudinal T2 hyperintensity affecting the middle blade of the corpus callosum while sparing the inner and outer blades. B, Axial T2-weighted images of 2 patients. B1, Symmetric involvement of the striatum in a patient with *NDUFAF5* variants. B2, Symmetric hyperintensity in the thalami in a patient with *NDUFAF5* variants. C1, FLAIR image of a patient with *LYRM7* variants shows small cysts in the periventricular and deep cerebral white matter. C2, T2-weighted image shows the well-defined border of cysts in a different patient with *LYRM7* variants. D1, Axial diffusion trace images show extensive diffusion restriction at the edge of the lesions in a patient with *BOLA3* variants. D2, Postcontrast T1-weighted image shows subtle enhancement at the edge of cysts in the same patient as in C2. E1 and E2, Axial T2-weighted images. E1, Symmetric hyperintensity of the middle cerebellar peduncles and cerebellar white matter in a patient with *ISCA2* variants. E2, Hyperintensity of the medulla oblongata, middle cerebellar peduncles, and cerebellar white matter in a patient with *IBA57* variants.

together with the remaining genetic subgroups, so, patients were clustered in 5 groups: complex 2; complexes 3 and 4; *NUBPL*; *KSS*; and other.

To identify which MR imaging features correlated with the 5 subgroups, we performed hierarchical testing. First, the association of each feature with the overall group structure was tested. Then, for each significant feature, the association with each of the 5 subgroups was tested. Fisher exact tests were used with Holm-Bonferroni correction for multiple testing to control the maximum family-wise error rate at 0.05.

RESULTS

MR Imaging Findings

The age of patients at the time of their first available MR image ranged from the first month of life to 46 years of age.

T2 hyperintensity affecting the middle blade of the corpus callosum, while sparing thin inner and outer blades, was noted in 85 of our 132 patients (Fig 1). Brain stem lesions were found in 66 patients (Fig 1). These included selective involvement of certain tracts, such as part of the corticospinal tracts in 32 patients but also more diffuse or ill-defined brain stem lesions. Symmetric T2 hyperintensity in the middle cerebellar peduncles was found in 42 patients (Fig 1). Rarefaction of white matter lesions was found in 50 patients; cystic lesions were found in an additional 20 patients. These cysts were typically well-delineated (Fig 1). Symmetric abnormalities of deep gray matter structures were found in 42 patients (Fig 1), more specifically in the thalamus in 34,

substantia nigra in 13, globus pallidus in 9, putamen in 8, and caudate nucleus in 5. Cerebellar atrophy was found in 23 patients. Multifocal enhancement was found in 31 of 66 patients with contrast-enhanced images, most frequently at the edges of lesions (Fig 1). Multifocal diffusion restriction was found in 68 of 84 patients with diffusion-weighted images, again, most frequently at the lesion edges (Fig 1). Elevated lactate levels were found in 31 of 49 patients with MR spectroscopy.

Brain stem lesions were, in particular, common in succinate dehydrogenase (SDH) deficiency (19 of 22 patients) and *KSS* (all 6 patients). Symmetric T2 hyperintensity in the middle cerebellar peduncles was more commonly seen in SDH deficiency (14 of 22 patients) and *KSS* (5 of 6 patients), but also in patients with *NDUFS1* variants (5 of 10 patients). Cystic lesions were most commonly observed in patients with *APOPT1* variants (5 of 7 patients), patients with *LYRM7* variants (6 of 10 patients), and also in patients with both *COX6B* and *COX10* variants.

We investigated whether certain MR imaging abnormalities or combinations of abnormalities were specific for particular disorders or clusters of disorders. Complex 2 deficiency was characterized by T2 hyperintensity in the brain stem, more specifically in the transverse pontine fibers, and T2-hyperintense abnormality in the middle cerebellar peduncles and the thalami (Fig 2). Defects in complexes 3 and 4 were characterized by predominantly periventricular localization of T2 hyperintensities and well-defined cystic lesions with distinct borders (Fig 3). Patients with *NUBPL* variants had a unique

combination of T2-hyperintense signal of the cerebellar cortex bilaterally and supratentorial white matter abnormalities (Fig 4). Patients with KSS had T2-hyperintense abnormalities predominantly affecting the directly subcortical cerebral white matter, in combination with T2-hyperintense abnormalities in the globus pallidus and substantia nigra (Fig 5).

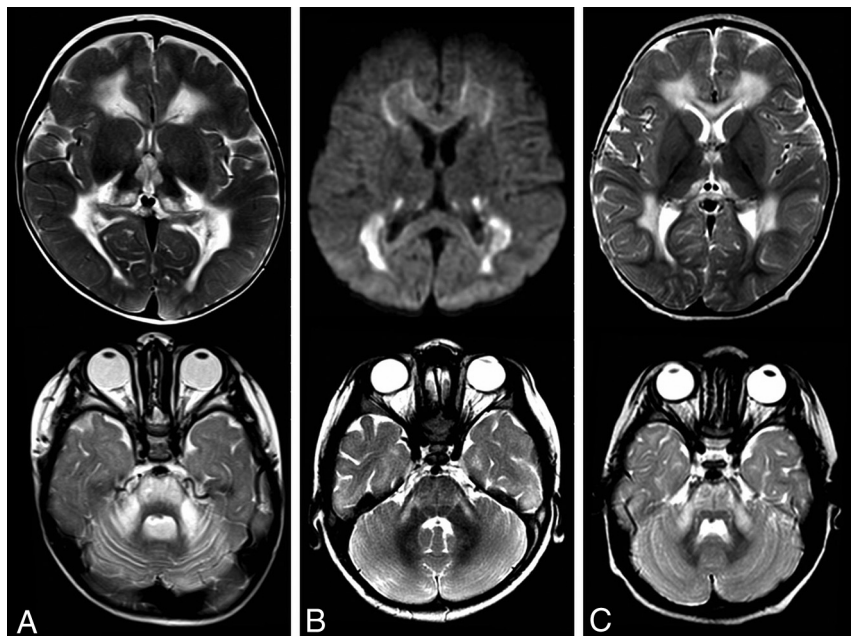


FIG 2. Features characteristic of SDH deficiency in 3 patients (A–C) showing symmetric thalamic involvement and pontine and middle cerebellar hyperintensities on T2-weighted images. B, Note the diffusion restriction.

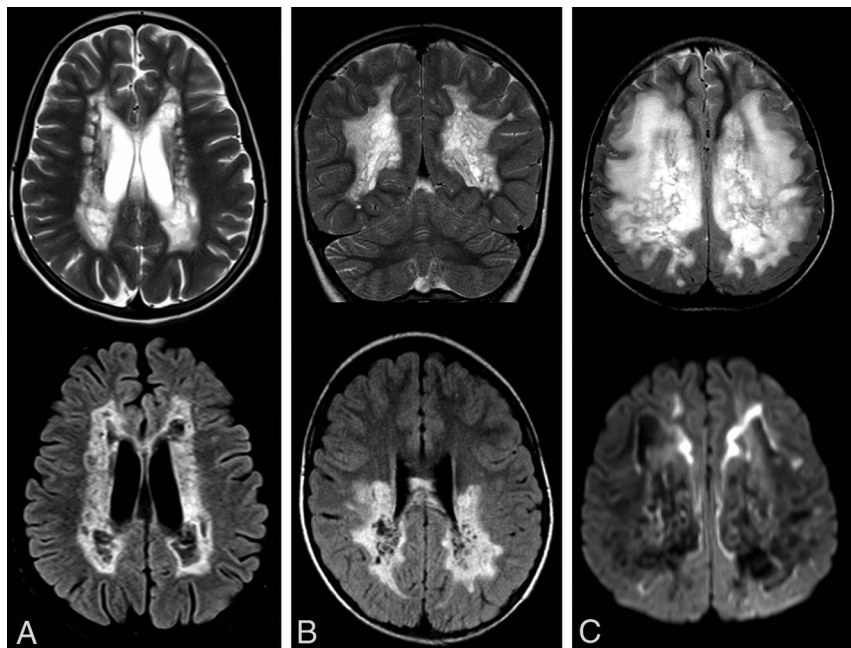


FIG 3. Characteristic features of defects in complexes 3 (A) and 4 (B and C). Upper rows are T2-weighted images; lower rows show FLAIR images and a diffusion trace image. Predominantly periventricular T2 hyperintensities, well-defined cystic lesions with a distinct border. Diffusion restriction at the edge of the lesions.

DISCUSSION

We investigated the MRIs of patients with leukodystrophies caused by a genetic defect affecting the mitochondrial respiratory chain function, except for mitochondrial transfer RNA synthetase defects. Several MR imaging features were relatively frequent among all patients and therefore suggestive of a possible mitochondrial cause when observed. Additionally, we found 4 MR imaging patterns specific for the following defects: 1) SDH deficiency, 2) *LYRM7* or *APOPT1* or *COX10* or *COX6B* variants, 3) *NUBPL* variants, and 4) KSS.

MR Imaging Features Suggestive of a Mitochondrial Leukodystrophy

In the diagnostic work-up of patients with leukodystrophy, several MR imaging features suggest an underlying mitochondrial defect (Table 1 and illustrated in Fig 1).

We observed longitudinal T2 hyperintensity of the middle blade of the corpus callosum in most of our patients, with sparing of inner and outer blades. In metachromatic leukodystrophy, Krabbe disease, and adrenoleukodystrophy, all layers of the corpus callosum are typically affected,²³ while in vanishing white matter, the inner blade is involved.²⁴ Marchiafava-Bignami disease²³ and Susac syndrome²⁵ also selectively affect the middle blade of the corpus callosum, in Marchiafava-Bignami disease in a longitudinal fashion; but in Susac syndrome, the lesions are round and multifocal.²⁵ Thus, longitudinal selective involvement of the middle layer should be regarded as a red flag for possible mitochondrial disease.

White matter rarefaction and cystic degeneration, most accurately visualized in FLAIR sequences, were common in our patients. The cysts were usually multifocal and well-delineated and mainly located in the periventricular and deep white matter and not in the subcortical regions. Diffusion restriction and contrast enhancement often specifically involved the rims of the cysts. By contrast, in Alexander disease, cysts are mainly located in the deep frontal white matter and may become quite large.²³ In megalencephalic leukoencephalopathy with subcortical cysts, the cysts are

located in the directly subcortical white matter.²³ Seldom, patients with Canavan disease may exhibit multiple small cysts throughout the white matter.²⁶ Bilateral anterior temporal lobe localization of the cysts, often seen in megalencephalic leukoencephalopathy with subcortical cysts, congenital cytomegalovirus infection, and Aicardi-Goutières syndrome,²³ was not observed in our patient cohort,

though this localization has been reported in a mitochondrial disorder caused by *RMND1* variants.²⁷ In vanishing white matter, the cerebral white matter rarefaction and cystic degeneration are diffuse and without well-defined rims.^{23,24} Therefore, the appearance and location of the white matter cysts are indicative of the type of leukodystrophy.

Symmetric deep gray matter involvement was common in our patients, the thalamus being most frequently involved. In Wilson disease, white matter abnormalities are uncommon but may be extensive in exceptional patients.²⁸ In Alexander disease, signal changes and mild swelling, followed by atrophy, occur in the caudate nucleus and putamen.²³ The globus pallidus and thalamus are commonly involved in Canavan disease with sparing of the putamen and caudate nucleus.²³ Other defects in amino acid and organic acid metabolism may present with white and deep gray matter abnormalities,²³ as well as in certain types of neurodegeneration with brain iron accumulation.²⁹ Therefore, the combination of white matter abnormalities and changes in deep gray matter nuclei comes with a long differential diagnosis, of which mitochondrial defects should be part.

Brain stem abnormalities were seen in half of our patients, involving both brain stem tracts and the pontine nuclei. The brain stem abnormalities in leukoencephalopathy with brain stem and spinal cord involvement and high lactate are distinct, involving the pyramidal tracts, medial lemniscus, cerebellar connections, and intraparenchymal trigeminal nerves.¹⁵ In peroxisomal disorders, brain stem abnormalities commonly occur together with abnormal signal of the cerebellar and parieto-occipital or frontal white matter.²³ In Wilson disease, abnormalities frequently diffusely affect the pontine tegmentum and midbrain tectum and tegmentum, creating the so-called “face of the giant panda.”³⁰ Diffuse or multifocal enhancing brain stem lesions occur in Alexander disease, particularly in the medulla and midbrain.²³ In dentatorubral-pallidoluysian atrophy, T2 hyperintensity exists in the midbrain and pons and often also in the globus pallidus and thalamus.²³ In adult patients, adult polyglucosan body

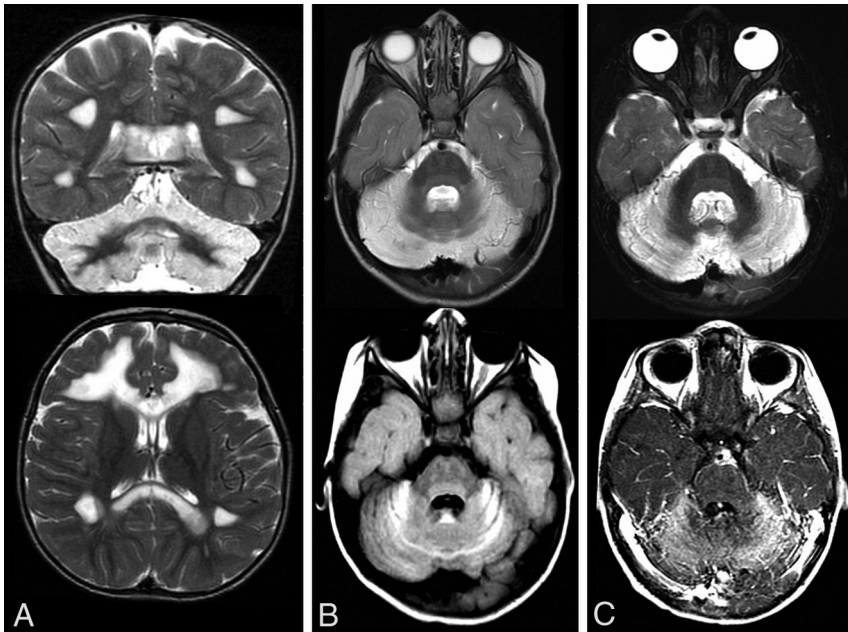


FIG 4. Characteristic features of *NUBPL* variants shown in 3 patients (A–C). The T2-weighted images show hyperintensity of the cerebellum, which is also hyperintense on FLAIR (B) and enhances (C). Involvement of supratentorial white matter can be extensive (A).

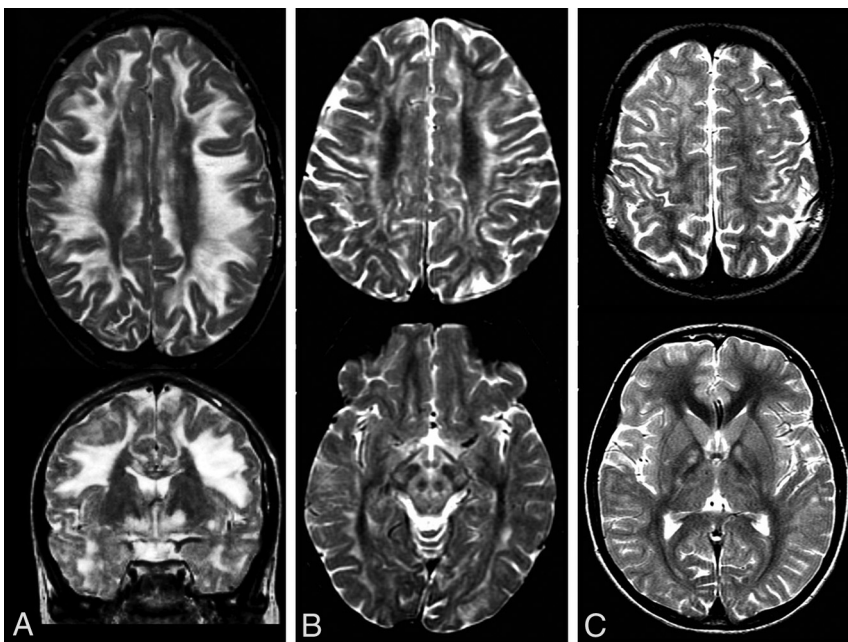


FIG 5. T2-weighted images of 3 patients with KSS (A–C) showing hyperintensity affecting (predominantly the subcortical and also deep) white matter. A and C, In the lower panel, hyperintensities in the globus pallidus exist. B, Involvement of the substantia nigra and, in this case, larger areas of the midbrain are shown.

Table 1: MR imaging features suggestive of a mitochondrial leukodystrophy

Imaging Features
Selective longitudinal involvement of the middle blade of the corpus callosum
Cerebral white matter rarefaction and cysts with a well-delineated rim, which may show contrast enhancement and diffusion restriction
Symmetric deep gray matter abnormalities
Brain stem abnormalities
Symmetric abnormalities in the middle cerebellar peduncles

Table 2: Diagnostic MR imaging patterns in mitochondrial leukodystrophies

MR Imaging Patterns
Succinate dehydrogenase (complex 2) deficiency
Abnormalities in the brain stem, more specifically in the transverse pontine fibers
Abnormalities in the middle cerebellar peduncles
Abnormalities in the thalami
Succinate in MR spectroscopy
Complex 3 and 4 defects
Numerous small cysts with well-defined borders in the periventricular and deep cerebral white matter
Kearns-Sayre syndrome
Abnormalities predominantly affecting the directly subcortical cerebral white matter plus abnormalities in the globus pallidus and substantia nigra
<i>NUBPL</i> defects
T2 hyperintensity of the cerebellar cortex plus cerebral white matter abnormalities

disease and autosomal dominant leukodystrophy with autonomic disease may be considered.^{31,32}

Symmetric T2 hyperintensity in the middle cerebellar peduncles was frequently seen in our patients, especially in SDH deficiency and KSS, and in those with *NDUFS1* variants. It has also been described in Wilson disease and adrenoleukodystrophy.³³ In adult patients with symmetric middle cerebellar peduncle involvement, autosomal dominant leukodystrophy with autonomic disease,³² fragile X-associated tremor/ataxia syndrome,²³ and multiple system atrophy can be considered.^{33,34}

Diagnostic MR Imaging Patterns

In some cases, the MR imaging pattern is specific for or strongly suggestive of a particular disorder or group of disorders and therefore quite helpful in the diagnosis (Table 2).

We found that patients with SDH deficiency could be diagnosed by the following MR imaging features: T2 hyperintensity in the brain stem (more specifically the transverse pontine fibers), the middle cerebellar peduncles, and thalami, as reported previously.¹⁹ Obviously, a succinate peak in MR spectroscopy is specific for SDH deficiency,¹⁹ but spectroscopy is not always available.

The presence of numerous small cystic lesions with a well-defined border in the periventricular and deep cerebral white matter was associated with complex 3 or 4 defects. In line with this association, the MR imaging findings previously described in patients with *LYRM7* variants¹⁴ are similar to those described in patients with *APOPT1* variants.¹² There are no previous descriptions of MR imaging abnormalities in patients with *COX10* or *COX6B* variants.

The combination of FLAIR or T2 hyperintensity of the cerebellar cortex and cerebral white matter involvement is unique for *NUBPL* variants.¹¹ Cerebellar cortical T2 hyperintensity has

been infrequently reported in other mitochondrial diseases,³⁵ but without cerebral white matter abnormalities. In several other diseases known for cerebellar cortical hyperintensity and volume loss,³⁶ typically no leukoencephalopathy exists.

A pattern of T2-hyperintense abnormalities predominantly affecting the U-fibers, the globus pallidus, and the substantia nigra was associated with KSS. Calcifications in the globus pallidus and substantia nigra may be visible. Brain stem and symmetric middle cerebellar peduncle abnormalities were also frequently observed in patients with KSS. However, these 2 features did not survive our statistical analysis, probably due to the small number of patients with KSS and the presence of these abnormalities in patients with SDH deficiency.

Study Limitations

A limitation of our study is its retrospective nature. Most important, this

limited the availability of important MR images, such as spectroscopy and contrast-enhanced images. Furthermore, a larger patient group would have increased the statistical power and may have led to the distinction of a higher number of specific MR imaging patterns. A strength of our study is that in all our patients, the causative gene defect was known. Still, it is difficult to estimate the generalizability of our results for new patients, though it is reassuring that similar, though limited, observations have been published before.^{11-14,19}

CONCLUSIONS

In patients with leukodystrophy, MR imaging findings are very helpful in directing the diagnostic process. In some cases, the pattern of MR imaging abnormalities is diagnostic, facilitating rapid genetic confirmation. This finding is also true for some mitochondrial leukodystrophies. In many of our patients, the MR imaging pattern was not specific for a particular mitochondrial defect, also in these patients MR imaging abnormalities suggestive of mitochondrial disease were commonly present.

ACKNOWLEDGMENTS

We thank the numerous colleagues who sent us MR imaging scans of their patients for review and provided us with the definitive diagnosis.

Disclosures: Adeline Vanderver—UNRELATED: Board Membership: Biogen, Takeda Pharmaceutical Company, Homology Medicines, Orchard Therapeutics, European Leukodystrophy Association (ELA) Foundation, United Leukodystrophy Foundation (ULF), Yaya Foundation, Metachromatic Leukodystrophy (MLD) Foundation, Ionis Pharmaceuticals, Passage Bio*; Grants/Grants Pending: National Institutes of Health, Takeda Pharmaceutical Company, Homology Medicines, Eli Lilly, Biogen, Ionis Pharmaceuticals, Pelizaeus Merzbacher Disease (PMD) Foundation, Foundation to Fight to Hypomyelination with Atrophy of the Basal Ganglia (H-ABC)*; Patents (Planned, Pending or Issued): Aicardi-Goutieres Syndrome (AGS)

scale, Hypomyelination with Atrophy of the Basal Ganglia (H-ABC) Therapeutics.* Nicole I. Wolf—UNRELATED: Consultancy: Shire/Takeda Pharmaceutical Company, Orchard Therapeutics, Ionis Pharmaceuticals, Passage Bio*; Grants/Grants Pending: ZonMW*; Travel/Accommodations/Meeting Expenses Unrelated to Activities Listed: European Society of Magnetic Resonance in Neuropediatrics. Marjo van der Knaap—UNRELATED: Grants/Grants Pending: ZonMw, European Leukodystrophy Association (ELA) Foundation, Hersenstichting, Vanishing White Matter Foundation, Chloe Saxby and Vanishing White Matter Disease Incorporated, and Vanishing White Matter Families Foundation Inc. Comments: ZonMw Top grant 91217006, European Leukodystrophy Foundation grant 2017-02712, European Leukodystrophy Foundation grant 2019-P001, Hersenstichting grant DR-2019-00285, Vanishing White Matter Foundation, Chloe Saxby and Vanishing White Matter Disease Incorporated, and Vanishing White Matter Families Foundation Inc; Patents (Planned, Pending or Issued): I have a patent, P112686CA00, on the therapeutic effects of Guanabenz treatment in vanishing white matter, pending to VU University Medical Center. *Money paid to the institution.

REFERENCES

- van der Knaap MS, Schiffmann R, Mochel F, et al. **Diagnosis, prognosis, and treatment of leukodystrophies.** *Lancet Neurol* 2019;18:962–72 [CrossRef Medline](#)
- Köhler W, Curiel J, Vanderver A. **Adulthood leukodystrophies.** *Nat Rev Neurol* 2018;14:94–105 [CrossRef Medline](#)
- Stellitano LA, Winstone AM, van der Knaap MS, et al. **Leukodystrophies and genetic leukoencephalopathies in childhood.** *Dev Med Child Neurol* 2016;58:680–89 [CrossRef Medline](#)
- Bonkowski JL, Nelson C, Kingston JL, et al. **The burden of inherited leukodystrophies in children.** *Neurology* 2010;75:718–25 [CrossRef Medline](#)
- Area-Gomez E, Schon EA. **Mitochondrial genetics and disease.** *J Child Neurol* 2014;29:1208–15 [CrossRef Medline](#)
- Baertling F, Klee D, Haack TB, et al. **The many faces of paediatric mitochondrial disease on neuroimaging.** *Childs Nerv Syst* 2016;32:2077–83 [CrossRef Medline](#)
- Morató L, Bertini E, Verrigni D, et al. **Mitochondrial dysfunction in CNS white matter disorders.** *Glia* 2014;62:1878–94 [CrossRef Medline](#)
- Chu BC, Terae S, Takahashi C, et al. **MRI of the brain in Kearns-Sayre syndrome.** *Neuroradiology* 1999;41:759–64 [CrossRef Medline](#)
- Millar WS, Lignelli A, Hirano M. **MRI of five patients with mitochondrial neurogastrointestinal encephalomyopathy.** *AJR Am J Roentgenol* 2004;182:1537–41 [CrossRef Medline](#)
- Kevelam S, Steenweg M, Srivastava S, et al. **Update on leukodystrophies: historical perspective and adapted definition.** *Neuropediatrics* 2016;47:349–54 [CrossRef Medline](#)
- Kevelam SH, Rodenburg RJ, Wolf NI, et al. **NUBPL mutations in patients with complex I deficiency and a distinct MRI pattern.** *Neurology* 2013;80:1577–83 [CrossRef Medline](#)
- Melchionda L, Haack TB, Hardy S, et al. **Mutations in APOPT1, encoding a mitochondrial protein, cause cavitating leukoencephalopathy with cytochrome c oxidase deficiency.** *Am J Hum Genet* 2014;95:315–25 [CrossRef Medline](#)
- Theunissen TE, Szklarczyk R, Gerards M, et al. **Specific MRI abnormalities reveal severe Perrault syndrome due to CLPP defects.** *Front Neurol* 2016;7:203 [CrossRef Medline](#)
- Dallabona C, Abbink TE, Carozzo R, et al. **LYRM7 mutations cause a multifocal cavitating leukoencephalopathy with distinct MRI appearance.** *Brain* 2016;139:782–94 [CrossRef Medline](#)
- van Berge L, Hamilton EM, Linnankivi T, et al. **LBSL Research Group. Leukoencephalopathy with brainstem and spinal cord involvement and lactate elevation.** *Brain* 2014;137:1019–29 [CrossRef Medline](#)
- van der Knaap MS, Bugiani M, Mendes MI, et al. **Biallelic variants in LARS2 and KARS cause deafness and (ovario)leukodystrophy.** *Neurology* 2019;92:e1225–37 [CrossRef Medline](#)
- Steenweg ME, Ghezzi D, Haack T, et al. **Leukoencephalopathy with thalamus and brainstem involvement and high lactate ‘LTBL’ caused by EARS2 mutations.** *Brain* 2012;135:1387–94 [CrossRef Medline](#)
- Dallabona C, Diodato D, Kevelam SH, et al. **Novel (ovario)leukodystrophy related to AARS2 mutations.** *Neurology* 2014;82:2063–71 [CrossRef Medline](#)
- Helman G, Caldovic L, Whitehead MT, et al. **SDH Study Group. MRI spectrum of succinate dehydrogenase-related infantile leukoencephalopathy.** *Ann Neurol* 2016;79:379–86 [CrossRef Medline](#)
- Torraco A, Ardisson A, Invernizzi F, et al. **Novel mutations in IBA57 are associated with leukodystrophy and variable clinical phenotypes.** *J Neurol* 2017;264:102–11 [CrossRef Medline](#)
- Gurgel-Giannetti J, Lynch DS, Paiva A. D, et al. **Novel complex neurological phenotype due to a homozygous mutation in FDX2.** *Brain* 2018;141:2289–98 [CrossRef Medline](#)
- van der Knaap MS, Breiter SN, Naidu S, et al. **Defining and categorizing leukoencephalopathies of unknown origin: MR imaging approach.** *Radiology* 1999;213:121–33 [CrossRef Medline](#)
- van der Knaap MS, Valk J. **Magnetic Resonance of Myelination and Myelin Disorders.** Springer-Verlag; 2005
- van der Lei H, Steenweg M, Barkhof F, et al. **Characteristics of early MRI in vanishing white matter.** *Neuropediatrics* 2012;43:22–26 [CrossRef Medline](#)
- Bourekas EC, Varakis K, Bruns D, et al. **Lesions of the corpus callosum: MRI and differential considerations in adults and children.** *AJR Am J Roentgenol* 2002;179:251–57 [CrossRef Medline](#)
- Drenckhahn A, Schuelke M, Knierim E. **Leukodystrophy with multiple beaded periventricular cysts in Canavan disease.** *J Inher Metab Dis* 2015;38:983–84 [CrossRef Medline](#)
- Ulrick N, Goldstein A, Simons C, et al. **RMND1-related leukoencephalopathy with temporal lobe cysts and hearing loss.** *Pediatr Neurol* 2017;66:59–62 [CrossRef Medline](#)
- Trocenlo JM, Woimant F, El Balkhi S, et al. **Extensive striatal, cortical, and white matter brain MRI abnormalities in Wilson disease.** *Neurology* 2013;81:1557–57 [CrossRef Medline](#)
- Kruer MC, Boddaert N, Schneider A, et al. **Neuroimaging features of neurodegeneration with brain iron accumulation.** *AJNR Am J Neuroradiol* 2012;33:407–14 [CrossRef Medline](#)
- Hitoshi S, Iwata M, Yoshikawa K. **Mid-brain pathology of Wilson’s disease.** *J Neurol Neurosurg Psychiatry* 1991;54:624–26 [CrossRef Medline](#)
- Mochel F, Schiffmann R, Steenweg ME, et al. **Adult polyglucosan body disease: natural history and key MRI findings.** *Ann Neurol* 2012;72:433–41 [CrossRef Medline](#)
- Melberg A, Hallberg L, Kalimo H, et al. **MR characteristics and neuropathology in adult-onset autosomal dominant leukodystrophy with autonomic symptoms.** *AJNR Am J Neuroradiol* 2006;27:904–11 [Medline](#)
- Okamoto K, Tokiguchi S, Furusawa T, et al. **MR features of diseases involving bilateral middle cerebellar peduncles.** *AJNR Am J Neuroradiol* 2003;24:1946–54 [Medline](#)
- Shrivastava A. **The hot cross bun sign.** *Radiology* 2007;245:606–07 [CrossRef Medline](#)
- Tan AP, Robles C, Mankad K. **Cerebellar atrophy with T2/FLAIR hyperintense cerebellar cortex.** *Childs Nerv Syst* 2018;34:601–03 [CrossRef Medline](#)
- Poretti A, Wolf N, Boltshauser E. **Differential diagnosis of cerebellar atrophy in childhood: an update.** *Neuropediatrics* 2015;46:359–70 [CrossRef Medline](#)